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Assessment of Heavy Metal Pollution in Water and Sediment of River Thiba, Kirinyaga County, Kenya

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ABSTRACT

Trace elements find their way into humans through ingestion, direct absorption or inhalation. All trace metals are toxic to animals and plants when present in excess amounts. The harmful effects of trace heavy metals in mammals may manifest as growth retardation, decrease in longevity, changes in reproductive cycles, chronic diseases and tumour formation. River Thiba catchment is Mt. Kenya forest and then flows through rocks, soil, farmlands, residential areas and town centres. Due to geological factors and human activities, trace heavy metals may be getting in river Thiba and since the water is used for domestic purposes and irrigation, it was necessary to determine the heavy metal concentration in the river. Sediment and water samples were collected along river Thiba during rainy and dry seasons. The samples were digested then concentrations of eight heavy metals determined using ICP-MS. The mean amounts of Cd, Cr, Ni, Pb, Zn, As, Mn and Se in sediment were 0.0908, 39.8969, 85.1085, 10.3918, 42.5555, 2.3679, 1678.3876 and 5.4907 mg/kg respectively during rainy season and 0.0628, 42.6319, 396.4692, 3.2669, 58.7585, 2.8139, 1766.4009 and 6.1059 mg/kg respectively during dry season. The mean concentration of the same metals in water was 0.0002, 0.0916, 1.0066, 0.0434, BDL, 0.0025, 1.8484 and 0.0038 ppm respectively during wet season and BDL, BDL, BDL, 0.0068, BDL, 0.00002, 0.0070 and BDL ppm respectively during dry season. The mean amount of Cr, Ni and Mn in sediment were found to be above WHO and US EPA permissible limits during both seasons. During rainy season, the mean concentration of Cr, Ni, Pb and Mn in water were above WHO and KEBS/WASREB permissible limits but the concentrations of Cd, Zn, As and Se were below the limits. However, during dry season, all the eight heavy metal concentrations in water were below WHO, KEBS/WASREB permissible limits. As far as the eight heavy metals are concerned, water from river Thiba may not be potable during rainy season but is potable during dry season. However, further investigations should be carried out to determine other water parameters.

Key words: Heavy Metals, Sediment Pollution, Water Pollution, Potable Water, Carcinogenic, Teratogenic.

1.0 INTRODUCTION

Heavy metals find their way into human body by ingestion, direct absorption or by inhalation. An indispensable link in the food chain is the terrestrial or marine plant life from which humans receive trace elements directly or indirectly by feeding on herbivorous animals which depend on plants for their nutrition. Plants absorb trace elements either via the root system or by foliar absorption [1]. Some of the heavy metals in traces are considered to be essential for plants and animals and they serve some useful biological functions [2]. Some of the heavy metals that can get in soil and water as a result of natural or human activities include Cd, Cr, Ni, Pb, Zn, As, Mn and Se.

Cadmium and zinc are usually found together in nature due to their similarities in atomic structure and chemical properties. Whereas zinc is an essential element in human body, cadmium is considered as a toxic element [1]. It has deleterious effects on bone structure and kidneys [3] and acts as inhibitor of sulphydryl enzymes. The symptoms of cadmium toxicity include hypertension, respiratory disorders, damage to kidneys and liver, aminoaciduria (urinary excretion of aminoacids), hypecalciurea (urinary excretion of excessive calcium), glycosuria (excretion of blood sugar

in the urine), proteinuria (urinary excretion of proteins), osteopororis (decalcification of the skeleton), formation of kidney stones and others [1, 4, 5, 6].

Chromium improves insulin sensitivity and also enhances carbohydrates, lipids and protein metabolism. Deficiency of chromium includes impaired glucose tolerance and less efficient control of cholesterol [7, 8]. Chromium toxicity depends on it's oxidation state with Cr (VI) being much more toxic than Cr (III). Actually, Cr (VI) is more easily absorbed than Cr (III) through oral or inhalation routes [9, 10]. Ingestion of Cr (VI) in high doses have resulted to acute effects on cardiovascular, respiratory, hepatic, haematological and even death [11, 12]. Chronic exposure to Cr (III) has been found to result in liver malfunction, renal failure, anaemia and weight loss [11, 12, 13].

Nickel is essential in animals and has been shown to be present in RNA and DNA [14]. It is essential to some microorganisms, animals (but not plants) and is associated with synthesis of vitamin B-12 although it is toxic at higher concentrations [1]. It is believed to inhibit various enzymes such as cytochrome oxidase, isocitrate dehydrogenase and maleic dehydrogenase. Nickel dust is also believed to be carcinogenic [1, 14].

Lead is a general protoplasmic poison which is cumulative, slow-acting, subtle and like other heavy metals, it has high affinity for sulphur. It exerts most of its activities through sulphydryl inhibition and also interacts with carboxyl and phosphoryl groups [1]. The major biochemical effects of lead is it's interference with heme synthesis leading to hematological damage and also inhibition of utilization of iron in the body [3]. Inorganic lead is able to replace calcium in bones and accumulate there as a reservoir for long-term release later to soft tissues [1, 15]. The most severe clinical form of lead poisoning is brain damage which produces clumsiness, subtle changes in mental attitude, sluggishness, poor memory, inability to concentrate, restlessness and hyper irritability [3]. Lead is quite toxic to young children because they cannot excrete much lead; consequently, they retain high percentage of ingested lead than adults. Exposure to lead can stunt a child's intellectual, behavioural and physical development [16].

Zinc is essential in human body and it's requirements increases during pregnancy, lactating period and growth age period [17, 18]. Moderate deficiency of zinc leads to retarded growth, delayed puberty, male hopogonadism, rough skin, poor appetite, slow wound healing, taste abnormalities and mental lethargy [19, 20]. Severe cases of zinc deficiency are diarrhoea, weight loss, pustular dermatitis, emotional disorder and intercurrent infections [19]. Zinc is required in protein synthesis, DNA synthesis, cell division, testicular size and functions and hormonal functions of various endocrine organs [21, 22]. Acute toxicity of zinc leads to nausea, gastric distress and disorientation [23]. Just like zinc deficiency, elevated zinc levels may compromise immunity [20, 21,24]. It has been found that if there is persistence of zinc intoxication, both copper and iron absorption are interfered with which may cause anaemia as a result of iron deficiency [25, 26].

Arsenic is a general protoplasmic poison and it affects all systems in the body. Major biochemical effects of arsenic are complexation with co-enzymes, uncoupling of phosphorylation and coagulation of proteins. Trivalent arsenicals combine with sulphydryl (-SH) groups in cells and thus inhibit the sulphydryl containing enzyme systems essential to cellular metabolism [27]. Low chronic doses of arsenic tend to accumulate in lipid rich tissues [28]. Arsenic is toxic to liver and it produces fatty infiltration and causes central necrosis and cirrhosis; affects bone marrow and cellular elements of blood [1]; is a carcinogen and greatly increases the risk of bladder cancer [5]. Arsenic also crosses placental membrane and is teratogenic [3, 6].

Manganese is found in bones, pancreas, kidneys and liver in little amounts [29, 30]. It helps in formation of connective tissues, clotting factors, bones, sex hormones and is necessary in brain and nerve functions. It is also an antioxidant enzyme superoxide dismutase (SOD) component [31, 32, 33]. Low quantities of manganese in the body may lead to bone malformation, infertility, weakness and seizures. However, large amounts of manganese in diet may lead to elevated manganese in body tissues [34, 35]. Manganese presents neurotoxicity that is unique because it begins with early psychiatric abnormalities and ends up with parkinsonian syndrome [30, 33]. It is an inducer of pro-inflammatory situation in glial cells through producing interleukins and Tumor Necrosis Factor-Alpha [36, 37]. Manganese triggers

mitochondrial disruption by promoting influx of calcium leading to functional loss and terminal permeability of mitochondrial membrane [38].

Selenium is a component of amino acids found in active sites of enzymes in animals and is required in trace amounts [30]. At high amounts, selenium is toxic because it replaces sulphur in enzymes [39, 40]. In humans, selenium deficiency can lead to muscle necrosis, hypothyroidism, cardio-cerebrovascular, male infertility and also may increase risk of development of cancers of lung, skin, colon, esophagus and stomach [9, 41]. Selenium is associated with pathogenesis of type two diabetes melitus, liver damage, increased serum liver enzyme levels, higher hepatic insulin resistance, and high triglyceride levels [42, 43]. Chronic exposure to selenium compounds affects synthesis of thyroid hormone, impairment of natural killer cells activity, hepatotoxicity and also gastrointestinal disturbances [44, 45, 46]. Acute exposure selenium leads to dermatological effects which includes nail loss, hair loss and dermatitis [46]. Extreme cases of selenium poisoning can result to liver cirrhosis, pulmonary edema and death [47, 48].

River Thiba flows from Mount Kenya forest though rocks and soil. Flowing downstream, the water passes through agricultural lands, residential areas and urban centers. Weathering of rocks; surface runoffs from agricultural farms, residential areas and towns; and seepage of waste water to the river may be contributing to the water getting polluted with heavy metals as it flows downstream. The river water is consumed by people and animals as well as being used for irrigation. Therefore, the objective of this study was to determine the levels of heavy metals in water and sediment of river Thiba.

2.0 MATERIALS AND METHODS

Water and sediment samples were collected in triplicates during dry and rainy season along rivers Thiba upstream from Kiumbu (about one kilometre downstream from Ngomano; the confluence of river Thiba and Nyamindi) to Ndiara (near Kianyaga town). The sampling points along the river were Kiumbu which was coded as C2, Ngomano (at the confluence of river Thiba and Nyamindi) and coded as C1, Kakawa (T1), Ndindiruku (T2), Kiamanyeki (T3), Kithogondo (T4), Kangai (T5), Karii (T6), Kutus (T7) and Ndiara (T8). The water samples were collected at a depth of about one foot below water surface (to avoid scum collection) using grab sampler then stored in plastic bottles. Important details such as collection date and code were noted. The samples were preserved by adding two drops of concentrated nitric acid to each of them before storing below 4° C until they were digested [49, 50, 51].

Sediment samples were collected (at the same point and time with water samples) using auger sampler in triplicates. Stones and plant materials were removed by hands then samples were stored in plastic containers that had previously have been cleaned with 10% nitric acid and rinsed with distilled water. The samples were labelled appropriately as described in water sample collection procedure and transported to laboratory where they were dried in an oven at 105^o C. To obtain analytical samples of sediment, coning and quartering technique was done [52]. Thereafter, the samples were ground to fine powder then stored at a temperature below 4° C.

A mass of 0.5±0.025 g of each sediment sample was weighed out using Top pan Sartorius balance into an ultra-clean and dry inert polymeric microwave vessel. A volume of 9±0.5 ml of concentrated nitric acid was slowly added under fume extraction hood. After this, 4 ± 0.05 ml of perchloric acid was added slowly followed by 1 ± 0.05 ml of concentrated hydrochloric acid. The samples were left to react for 5 ± 1 minute prior to sealing the vessels to allow gases to escape. The vessels were placed on the rotor then placed in the microwave digester (High performance microwave digestion system, ETHOS UP). The microwave digester was gradually heated to 150 \degree C in over 20 minutes then held at that temperature for another 20 minutes. After the last 20 minutes, the microwave automatically began cooling the samples and indicated when cooling was complete. After the cooling, the samples were quantitatively transferred into 250 ml volumetric flasks then filled to the mark using distilled de-ionized water [53, 54, 55].

The water sample was shaken thoroughly in their plastic containers by use of hand. A volume of 45 ml of the sample was measured into an ultra-clean and dry inert polymeric microwave vessel. A volume of 9±0.5 ml of concentrated nitric acid was then slowly added under fume extraction hood followed by 4 ± 0.05 ml of perchloric acid. The vessels

were placed on the rotor then placed in the microwave digester (High performance microwave digestion system, ETHOS UP) then digestion and topping up to 250 ml was done in a similar way as was done for sediment.

Multi element standards of 10 ppm from Agilent Technologies was used as the stock solution. A calibrated micro pipette was used to transfer 0, 100, 200,300, 500 and 1000 µL of stock solution into well labelled 100 ml volumetric flasks to prepare 0,10, 20,30, 50 and 100 ppb standards, respectively. Each of these volumetric flasks was topped up to the marks using 5% nitric acid solution. These standards were used for preparing calibration curve. Heavy metal (Cd, Cr, Ni, Pb, Zn, As, Mn and Se) analysis was done in chemistry laboratories using inductively coupled plasma-mass spectrometry (Agilent Technologies 7900 ICP-MS). The data was analyzed using statistical package for social sciences (SPSS) Version 26.

3.0 RESULTS AND DISCUSSION

3.1 Heavy Metals in Sediment of River Thiba from Kiumbu to Ndiara During Rainy Season

The amount of selected heavy metals in sediment from river Thiba during rainy season are presented in Table1.

Table 1: Amount of Selected Heavy Metals (mg/kg) in Sediment Along River Thiba during Rainy Season

The amount of Cd in sediment from Kiumbu to Ndiara along river Thiba during rainy season ranged from 0.0240 \pm 0.0004 (T5) to 0.2705 \pm 0.0076 (T7) mg/kg with an average of 0.0908 \pm 0.0790 mg/kg. Cr values in the sediment ranged from 33.1244±0.0053 (T5) to 50.3350±0.0140 (T6) mg/kg and had a mean of 39.8969±5.7550 mg/kg. For Ni, values ranged from 67.8782 ± 0.0550 (T4) to 128.1840 ± 0.3980 (T6) mg/kg with a mean of 85.1085±18.0378 mg/kg. Pb values were ranging between 7.4881±0.0059 (T2) and 12.9476±0.0055 (T7) mg/kg with a mean of 10.3918±1.8690 mg/kg.

The amount of Zn in the sediment varied from 37.6893 ± 0.0049 (T1) to 50.5437 ± 0.0340 (T7) mg/kg and had an average of 42.5555±4.0501 mg/kg. The values of As were ranging from 1.9632±0.0076 (T4) to 3.1320±0.0071 (C1) mg/kg with a mean of 2.3679 ± 0.3390 mg/kg. For Mn, the amount varied from 1217.8688 ± 0.7570 (C2) to

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2193.8618±6.5110 (T3) mg/kg with a mean of 1678.3876±292.8288 mg/kg. Whereas Se values ranged from 3.7801 \pm 0.0025 (T6) to 7.1824 \pm 0.0046 (T5) mg/kg with an average of 5.4907 \pm 1.1195 mg/kg. The mean amounts of the heavy metals in sediment along river Thiba upstream from Kiumbu to Ndiara during wet season decreased in the order Mn>Ni>Zn>Cr>Pb>Se>As>Cd

These amounts of the heavy metals did not show any uniform general trend (increase or decrease) upstream from Kiumbu to Ndiara. It was noted that at T7 (Kutus), the values were highest for Cd, Pb and Zn and at T6 (Karii) which was the next sampling point after Kutus downstream had highest values of Cr and Ni. This could be due to T6 and T7 sampling points close proximity to Kutus town which points to a possibility of the heavy metals from this town finding their way to the river sediment through surface run-offs, seepage or both. During the rainy season, the mean amounts of Cd, Pb, Zn, As and Se in the sediment fell below the WHO and US EPA limit but the mean amounts of Cr, Ni and Mn were above the limits.

The present study obtained mean amount of Pb of 10.3918 ± 1.8690 mg/kg during wet season which is lower than 22.95±15.11 mg/kg that was reported by Gathumbi and collaborators [56]. This may be due to phasing out of leaded petrol which was earlier used in petrol powered water pumps that are used in Mwea region. Moywaywa reported higher mean amounts of Mn, Ni, Zn and Pb in sediment from Thika river [57] showing that Thika river may be more polluted with heavy metals than river Thiba. Omondi reported lower Mn concentration but higher Ni, Cr, Zn, Pb and As concentration in sediment from river Nzoia [58] which shows that river Thiba may be less polluted than river Nzoia. Idiriah and collaborators reported far much lower concentrations of Zn, Pb and Cr in sediment collected along Abonnema shoreline in Nigeria [59].

3.2 Heavy Metals in Sediment of River Thiba from Kiumbu to Ndiara During Dry Season

The amount of selected heavy metals in sediment sampled along river Thiba during dry season are given in Table 2.

Table 2: Amount of Selected Heavy Metals (mg/kg) in Sediment Along River Thiba during Dry Season

The amount of Cd in sediment during dry season varied from 0.0292 ± 0.0008 (C2) to 0.1242 ± 0.0027 (T2) mg/kg with an average of 0.0628±0.0276 mg/kg. Cr values ranged from 26.1350±0.0076 (T3) to 64.0501±0.0490 (T7) mg/kg with a mean of 42.6319±11.4988 mg/kg. Ni amount varied from 63.2156±0.0570 (T3) to 741.2449±0.6250 (T6) mg/kg with a mean of 396.4692±238.9054 mg/kg. Amount of Pb varied from 0.0523±0.0004 (T6) to 10.2168±0.0044 (T2) mg/kg with a mean of 3.2669±4.7021 mg/kg. Zn amounts ranged from 32.8121±0.0079 (T2) to 88.9347±0.0260 (T7) and had an average of 58.7585±19.5531 mg/kg. For As, the amount varied from 2.2627±0.0063 (T3) 3.4536±0.0064 (T6) mg/kg with an average of 2.8139±0.3721 mg/kg. Mn had amounts varying from 1597.5113±2.6710 (T2) 1862.6627±4.5260 (T5) mg/kg with a mean of 1766.4009±92.0517 mg/kg. Whereas Se values ranged from 4.0637±0.0066 (T3) 8.4542±0.0085 (T6) mg/kg with a mean of 6.1059±1.4419 mg/kg. The mean amounts of the heavy metals in the sediment during dry season decreased in the order Mn>Ni>Zn>Cr>Se>Pb>As>Cd. Just as during rainy season, it was noted that during dry season, sampling point T7 (Kutus) had highest amounts of Cr and Zn; T6 (Karii) which is the next sampling point after Kutus downstream had highest values of Ni, As and Se suggesting that there may be activities in and/or around Kutus town that contributeing to higher amounts of these heavy metals in the sediment. During dry season, the average content of Cd, Pb, Zn, As and Se in sediment were below the limit set by WHO and US EPA but the average contents of Cr, Ni and Mn were above the limits.

The current study obtained mean amount of Pb of 3.2669 \pm 4.7021 mg/kg during dry season which is much lower than 22.95±15.11 mg/kg that was reported by Gathumbi and collaborators on a study carried out in the same area [56]. This may be an indication that lower quantities of Pb may be finding their way in river Thiba since petrol powered water pumps that are usually placed next to the river use unleaded petrol. Moywaywa reported higher mean amounts of Mn, Zn and Pb in sediment from Thika river [57] implying that Thika river could have been more polluted with these heavy metals than Thiba river. Omondi [58] reported lower amount of Mn, close amount of Ni and higher amounts of Cr, Zn, Pb and As in sediment from river Nzoia showing that river Thiba sediment was less polluted than river Nzoia.

3.3 Heavy Metals in Water of River Thiba from Kiumbu to Ndiara During Rainy Season

The amounts of selected heavy metals in water sampled along river Thiba during rainy season are given in Table 3.

Point	Cd	Cr	Ni	Pb	Zn	As	Mn	Se
C ₂	$0.0003\pm$	$0.1536 \pm$	$1.9008 +$	0.0038 _±	BDL	$0.0074\pm$	$6.8320 \pm$	0.0092 _±
	0.00004	0.00015	0.00073	0.00006		0.00003	0.00052	0.00004
C1	$0.0004\pm$	$0.1768 +$	$0.8468 \pm$	0.0135±	BDL	$0.0014\pm$	1.3190±	BDL
	0.00003	0.00071	0.00031	0.00004		0.00008	0.00028	
T1	BDL	BDL	BDL	BDL	BDL	$0.0001\pm$	$0.0177 \pm$	BDL
						0.00001	0.00002	
T ₂	$0.0004\pm$	$0.1214 \pm$	$3.2534 \pm$	$0.0085\pm$	BDL	$0.0060 \pm$	$3.2295 \pm$	$0.0138 +$
	0.00002	0.00033	0.00062	0.00002		0.00003	0.00043	0.00003
T ₃	$0.0003\pm$	$0.1927 \pm$	$2.0554 \pm$	$0.0141 \pm$	BDL	0.0048 _±	$3.4972 \pm$	0.0092 _±
	0.00006	0.00024	0.00035	0.00005		0.00004	0.00035	0.00002
T ₄	$0.0001\pm$	$0.0960 \pm$	$1.1054 +$	0.0163 [±]	BDL	$0.0025 \pm$	$1.5181 +$	BDL
	0.00005	0.00004	0.00039	0.00007		0.00005	0.00022	
T ₅	BDL	BDL	$0.0701 \pm$	$0.0024 \pm$	BDL	0.0003 _±	$0.2879 \pm$	$0.0004\pm$
			0.00006	0.00001		0.00006	0.00014	0.00008
T ₆	$0.0002\pm$	$0.0561 \pm$	$0.4881 \pm$	$0.0244 \pm$	BDL	$0.0020 \pm$	$1.4682 +$	$0.0058 +$
	0.00001	0.00007	0.00053	0.00002		0.00005	0.00044	0.00007
T7	$0.0001\pm$	$0.1196 \pm$	$0.3455 \pm$	$0.0999\pm$	BDL	$0.0004\pm$	$0.2959 \pm$	BDL
	0.00001	0.00067	0.00044	0.00005		0.00003	0.00048	
T ₈	BDL	BDL	BDL	$0.1243 +$	BDL	$0.0001\pm$	$0.0183 +$	BDL
				0.00007		0.00008	0.00003	
Mean	$0.0002\pm$	$0.0916 \pm$	$1.0066 \pm$	$0.0434 \pm$	BDL	$0.0025 \pm$	$1.8484 \pm$	$0.0038 +$
	0.0002	0.0741	1.0849	0.0554		0.0027	2.1484	0.0052
Limit	0.005	0.05	0.1	0.015	5.0	0.01	0,1	0.01

Table 3: Amount of Selected Heavy Metals (ppm) in Water Along River Thiba during Rainy Season

The concentration of Cd in river Thiba water from Kiumbu (C2) to Ndiara (T8) during rainy season varied between BDL and 0.0004±0.00003 (C1) ppm and had a mean of 0.0002±0.0002 ppm. Cr concentration ranged from BDL to 0.1927 ± 0.00024 (T3) ppm with a mean of 0.0916 ± 0.0741 ppm. Nickel concentration varied from BDL to 3.2534±0.00062 (T2) ppm with an average of 1.0066±1.0849 ppm. Concentration of Pb ranged between BDL and 0.1243 ± 0.00007 (T8) ppm and had an average of 0.0434 ± 0.0554 ppm. The concentration of Zn in all the sampling points along river Thiba course was BDL. As concentration was ranging from 0.0001 ± 0.00008 (T8) ppm to 0.0074 ± 0.00003 (C2) ppm and had a mean of 0.0025 ± 0.0027 ppm. Mn had concentration varying from 0.0177 \pm 0.00002 (T1) ppm to 6.8320 \pm 0.00052 (C2) ppm and a mean of 1.8484 \pm 2.1484 ppm. Whereas the concentration of Se ranged between BDL and 0.0138±0.00003 (T2) ppm with a mean of 0.0038±0.0052 ppm. The mean concentrations of the heavy metals in water along river Thiba upstream from Kiumbu to Ndiara during wet season decreased in the order Mn>Ni>Cr>Pb>Se>As>Cd>Zn. It was observed that there was no trend in increase or decrease in concentrations of the heavy metals in water sampled from river Thiba water either downstream or upstream between Kiumbu and Ndiara during the rainy season.

The sampling point T2 (Ndindiruku) had the highest concentration of Ni and Se whereas sampling point C2 (Kiumbu) had the highest concentrations of As and Mn. It was noted that at sampling point T1 (Kakawa), the concentration of Cd, Cr, Ni, Pb, Zn and Se were BDL. Similarly, it was noted that the concentration of Zn in water from all the sampling points along river Thiba was BDL. During the rainy season, the mean concentrations of Cr, Ni, Pb and Mn in the river water were above WHO and KEBS/WASREB permissible limit but the mean concentrations of Cd, Zn, As and Se fell below the limits. This study obtained lower concentrations levels of Pb and Cd than the ones reported by Wasike and collaborators [60] on water of Kuywa river during wet season. [60] also reported lower level of Mn than what is reported in this study. Muiruri and collaborators [61] reported very close concentration levels of Pb and Cd, lower levels of Ni, Cr and Mn and higher level of Zn in river water from Athi-Galana-Sabaki tributaries during wet season compared to what is reported in the current study.

3.4 Heavy Metals in Water of River Thiba from Kiumbu to Ndiara During Dry Season

The concentration of selected heavy metals in water sampled along river Thiba during dry season are given in Table 4.

Point	Cd	Cr	Ni	Pb	Zn	As	Mn	Se
C ₂	BDL	BDL	BDL	$0.0186 \pm$	BDL	0.0000	$0.0067\pm$	BDL
				0.00005			0.00001	
C1	BDL	BDL	BDL	$0.0022 \pm$	BDL	BDL	$0.0061 \pm$	BDL
				0.00003			0.00003	
T1	BDL	BDL	BDL	$0.0011 \pm$	BDL	$0.0001\pm$	$0.0096\pm$	BDL
				0.00003		0.00002	0.00001	
T ₂	BDL	BDL	BDL	$0.0130 \pm$	BDL	BDL	$0.0040 \pm$	BDL
				0.00005			0.00003	
T ₃	BDL	BDL	BDL	0.0038 [±]	BDL	BDL	$0.0093\pm$	BDL
				0.00007			0.00004	
T4	BDL	BDL	BDL	0.0032 _±	BDL	0.0000	0.0037 _±	BDL
				0.00008			0.00006	
T ₅	BDL	BDL	BDL	$0.0077\pm$	BDL	0.0000	$0.0066 \pm$	BDL
				0.00006			0.00006	
T7	BDL	BDL	BDL	$0.0048 \pm$	BDL	$0.0001\pm$	$0.0083\pm$	0.0000
				0.00004		0.00008	0.00005	
T8	$0.0073\pm$	BDL	BDL	$0.0069\pm$	BDL	0.0000	$0.0085\pm$	BDL
	0.00005			0.00008			0.00008	
Mean	BDL	BDL	BDL	$0.0068 +$	BDL	$0.00002\pm$	$0.0070 \pm$	BDL
				0.0057		0.00004	0.0021	
Limit	0.005	0.05	0.1	0.015	5.0	0.01	0.1	0.01

Table 4: Amount of Selected Heavy Metals (ppm) in Water Along River Thiba during Dry Season

During the dry season, the concentration of cadmium in river Thiba water from Kiumbu to Ndiara was BDL for the all sampling points except T8 (Ndiara) where the concentration was 0.0073 ± 0.00005 ppm. The concentrations of chromium, nickel, zinc and selenium was also BDL for all the sampling points except selenium at T7 whose concentration was 0.0000 ppm. The concentration of lead was ranging between 0.0011 ± 0.00003 (T1) ppm and 0.0186 ± 0.00005 (C2) ppm and had an average of 0.0068 ± 0.0057 ppm. The concentration of arsenic was varying from BDL to 0.0001 ± 0.00002 (T1) ppm with a mean of 0.0068 ± 0.0057 ppm while the concentrations of manganese was ranging from 0.0037 ± 0.00006 (T4) ppm to 0.0096 ± 0.00001 (T1) ppm with a mean of 0.0070 ± 0.0021 ppm. The mean concentrations of the heavy metals in water along river Thiba upstream from Kiumbu to Ndiara during dry season decreased in the order Mn>Pb>Cd>As>Cr/Ni/Zn/Se. It was observed that most sampling points in the river had the heavy metal concentrations BDL apart from lead, manganese and arsenic (at some sampling points) which recorded low concentrations.

During the dry season, the mean concentrations of all eight heavy metals fell below WHO and KEBS/WASREB limit [16, 62] implying that the river water was not polluted with these metals. The study obtained lower concentrations levels of Pb, Cd and Mn than those reported on water of Kuywa river during dry season [60]. Muiruri and collaborators reported higher concentration levels of Ni, Mn, Zn, Cd and Cr in water from Athi-Galana-Sabaki tributaries during dry season [61] compared to findings of this study indication that river Thiba is less polluted compared to these rivers. The study [61] reported concentration of lead in the tributaries water that was close to what is reported in this study.

4.0 CONCLUSION AND RECOMMENDATIONS

Sediment from rivers Thiba contained all the eight heavy metals at varying levels. During both rainy and dry seasons, the mean amounts of Cd, Pb, Zn, As and Se were found to be below the permissible limit of WHO and US EPA for unpolluted sediment but the mean amounts of Cr, Ni and Mn were above the limits. During rainy season, the mean concentrations of Cd, Zn, As and Se in water were below the permissible limit set by WHO and KEBS/WASREB but Cr, Ni, Pb and Mn mean amounts were above the limits. However, during dry season, all the eight heavy metal mean amounts in water from the river were below WHO and KEBS/WASREB permissible limit which shows that water from this river may not be polluted with these heavy metals and so may be used as potable water (as far as the 8 heavy metals are concerned). It is recommended that people should avoid drinking water from river Thiba during rainy season and further studies require to be conducted to determine other water parameters (physical, biological and chemical) to confirm whether water from this river is potable.

REFERENCES

- 1. Dara S. S., & Mishra D. D. (2010). Environmental Chemistry and Pollution Control. S. Chand & company Ltd, New Delhi. Pp. 116-119, 159-184.
- 2. Tuorma, T. E. (2002). Chromium, Selenium, Copper and other Trace Minerals in Health and Reproduction. *The Journal of Orthomolecular Medicine*, 15, 145.
- 3. Jumba, I. I., & Likimani, T. A. (2001). Chemistry and its Application. Nairobi University Press, Nairobi. Pp. 170- 174, 249-257.
- 4. Qu, F. & Zheng, W. (2024). Cadmium Exposure: Mechanisms and Pathways of Toxicity and Implication for Human Health. *Toxics*, 12,388
- 5. Girard, J. E. (2014). Principles of Environmental Chemistry.3rded. Jones & Bartlett Learning, Burlington. Pp. 385- 438.
- 6. Duggal, K. N. (2007). Elements of Environmental Engineering. S. Chand and Company Ltd, Ram Nagar, New Delhi. Pp. 97.
- 7. Lewicki, S., Zdanowski, R., Krzzowska, M., Lewicka, A., Debski, B., Niemcewicz, M. & Goniewicz, M. (2014). The Role of Chromium III in the Organism and it's possible Use in Diabetes and Obesity Treatment. *Annals of Agricultural and environmental Medicine*, 21(2),331-335.
- 8. Pechova, A. & Pavlata, L. (2007). Chromium as an Essential Nutrient: A Review. *Veterinarni Medicina*, 52,1-18.

- 9. Genchi, G., Lauria, G., Catalano, A. & Sinicropi, M. S. (2021). The Double Face of Metals: The Intriguing Case of Chromium. *Applied Sciences*, 11(2),638.
- 10. Desmarais, T. L. & Costa, M. (2019). Mechanisms of Chromium Induced Toxicity. *Current Opinion on Toxicology*, 14,1-7.
- 11. Hessel, E. V. S., Staal, Y. C. M., Piersma, A. H., Braver-Sewrady, S. P. & Ezendam, J. (2021). Occupational Exposure to Hexavalent Chromium Part I. *Hazard Assessment of Non-Cancer Health Effects*, 126,105048
- 12. Ray, R. R. (2016). Adverse Hematological Effects of Hexavalent Chromium: An Overview. *Interdisciplinary Toxicology*, 9(2),55-65.
- 13. Balali-Mood, M., Naseri, K., Tahergorabi, Z., Khazdair, M. R. & Sadeghi, M. (2021). Toxic Mechanisms of Five Heavy Metals: Mercury, Lead, Chromium, Cadmium and Arsenic. *Frontiers in Pharmacology*, 12,643972
- 14. Samal, L. & Mishra, C. (2011). Significance of Nickel in Livestock and Production. *International Journal for Agro Veterinary and Medical Sciences*, 5,349-361
- 15. Wani, A. I., Ara, A. & Usmani, J. A. (2015). Lead Toxicity: A Review. *Interdisciplinary Toxicology*, 8(2), 55-64
- 16. World Health Organization (WHO) (2010). Childhood Lead Poisoning, WHO, Geneva, Switzerland. Pp 20-30.
- 17. Roohani, N., Hurrol, R., Kelihadi, R. & Schulin, R. (2013). Zinc and it's Importance for Human Health: An Integrative Review. *Journal of Research in Medical Sciences*, 18(2),142-157.
- 18. Donongelo, C. M. & King, J. C. (2012). Maternal Zinc Intakes and Homeostatic Adjustment During Pregnancy and Lactation, *Nutrients*, 4(7),782-798.
- 19. Prasad, A. S. (2013). Discovery of Human Zinc Deficiency: It's Impact on Human Health and Disease. *Advances in Nutrition*, 4(2),176-190.
- 20. Siklar, Z., Tuna, C., Dallar, Y. & Tanyer, G. (2003). Zinc Deficiency: A Contributing Factor in Short Stature in Growth Hormone Deficient Children. *Journal of Tropical Pediatrics*, 49(3), 187-8
- 21. Fallah, A., Mohammad-Hassani, A. & Colager, A. H. (2018). Zinc is an Essential Element for Male Fertility: A Review of Zinc Roles in Men's Health, Germination, Sperm Quality and Fertilization. *Journal of Reproduction Infertility*, 19(2), 69-81.
- 22. Wessels, I., Maywald, M. & Rink, L. (2017). Zinc as a Gatekeeper of Immune Function. *Nutrients*, 9(12),1286.
- 23. Fosmire, G. J. (1990). Zinc Toxicity. *American Journal of Clinical Nutrition*., 51(2),227-7.
- 24. Grungreiff, K., Gottstein, T. & Reinhold, D. (2020). Zinc Deficiency An Independent Risk Factor in the Pathogenesis of Haemorrhagic Stroke? *Nutrients*, 12(11), 3548.
- 25. Wahab, A., Mushtag, K., Borak, S.G. & Bellam, N. (2020). Zinc Induced Copper Deficiency, Sideroblastic Anaemia and Neutropenia: A Perplexing Facet of Zinc Excess. *Clinical Cases Reports*, 8(2),1666-1671.
- 26. Zahra, N., Kalim, I., Mahamood, M. & Naeem, N. (2017). Perilous Effects of Heavy Metals Contamination on Human Health. *Pakistan Journal of Analytical and Environmental Chemistry*, 18(1),1-17.
- 27. Thakur, M., Rachamalla, M., Niyogi, S., Datusolia, A. K. & Flora, J. S. (2021). Molecular Mechanism of Arsenic-Induced Neurotoxicity including Neuronal Dysfunctions, *International Journal of Molecular Science*, 22(18),10077
- 28. Jaishanker, M., Tseten, T., Abalagam, N., Mathew, B. B., & Beeregowda, K. N. (2014). Toxicity, Mechanism and Health Effects of Some Heavy Metals. *Interdisciplinary Toxicology*, 7(2), 60-72.
- 29. Soetan, K. O., Olaiya, C. O. & Oyewole, O. E. (2010). The Importance of Mineral Element for Humans, Domestic Animals and Plants: A review. *African Journal of Food Science*, 4(5),200-222.
- 30. Wada, O. (2004). What are Trace Elements?- Their Deficiency and Excess States. *Trace Elements*, 47(8),351-358.
- 31. Michalak, M., Pierzak, M., Krecisz, B. & Suliga, E. (2021). Bioactive Compounds for Skin Health: A Review. *Nutrients*, 13(1),203
- 32. Asif, M. (2017). Role of Heavy Metal in Health and Particularly in Respect to Diabetic Patients. *TANG* (*Humanitas Medicine*), 7(1), 1-10.
- 33. Avila, D. S., Puntel, R. L. & Aschner, M. (2013). Manganese in Health and Disease. *Metal Ions in Life Sciences*, 13,199-227.
- 34. Sriram, K. & Lonchyna, V. A. (2013). Micronutrient Supplementation in Adult Nutrition Therapy: Practical Considerations. *Journal of Parenteral and Enteral Nutrition*. 33(5),548-462
- 35. Thompson, L. J., Hall, J. O. & Meerdink, G. L. (1991). Toxic Effects of Trace Elements Excess. *Veterinary Clinics of North America: Food Animal Practice*, 7(1),277-306.

- 36. Kirkley, R. B., Popichak, K.D., Afzali, M. F., Legare, M. E. & Tjalkens, R. B. (2017). Microglia Amplify Inflammatory Activation of Astrocytes in Manganese Neurotoxicity. *Journal of Neuroinflammation*, 14,99
- 37. Jana, M., Dasgupta, S., Saha, R. N., Liu, X. & Paham, K. (2007). Introduction of Tumour Necrosis Factor-α (TNT-α) by Interleukin-12 P 40 Monomer and Homodimer in Microglia and Macrophages. *Journal of Neurochemistry*, 86(2),519-528.
- 38. Roa, K. V. R. & Norenberg, M. D. (2004). Manganese Induces the Mitochondrial Permeability Transition in Cultured Astrocytes. *Journal of Biological Chemistry*, 279(31),32333-8.
- 39. Hariharan, S. & Dharmaraj, S. (2020). Selenium and Selenoproteins: It's Role in Regulation of Inflammation. *Inflammopharmacology*, 28,667-695.
- 40. Maroney, M. J. & Hondal, R. J. (2018). Selenium versus Sulfur: Reversibility of Chemical Reactions and Resistance to Permanent Oxidation in Proteins and Nucleic Acids. *Free Radical Biology and Medicine*. 1(127),228-237.
- 41. Kieliszek, M., Bano, I. & Zare, H. (2011). A Comprehensive Review on Selenium and it's Effects on Human Health and Distribution in Middle Eastern Countries. *Biological Trace Elements Research*, 200,971-987.
- 42. Ogawa-Wong, A. N., Berry, M. J. & Seale, L. A. (2016). Selenium and Metabolic Disorders: An Emphasis on Type 2 Diabetes Risk. *Nutrients*, 8(2),80.
- 43. Yang, Z., Yan, C., Liu, G., Niu, Y., Zhang, W., Lu, S., Li, X., Zhang, H., Ning, G., Fan, J., Qin, L. & Su, Q. (2016). Plasma Selenium Levels and Nonalcoholic Fatty Liver Disease in Chinese Adults: A Cross-sectional Analysis. *Scientific Reports*, 6,37288.
- 44. Ventura, M., Melo, M. & Carrilho, F. (2017). Selenium and Thyroid Disease: From Pathophysiology to Treatment. *International Journal of Endocrinology* 2017,1297658
- 45. Kieliszek, M. & Blazejack S. (2016). Current Knowledge on Importance of Selenium in Food for Living Organisms: A Review. *Molecules*, 21(5),609.
- 46. Vinceti, M., Wei, E. T., Malagoli, C., Bergomi, M. & Vivoli, G. (2001). Adverse Health Effects of Selenium in Humans. *Reviews on Environmental Health*, 16(4),233-51.
- 47. Hadrup, N. & Ravn-Haren, G. (2020). Acute human Toxicity and Mortality After Selenium Ingestion: A Review. *Journal of Trace Elements in Medicine and Biology*, 5,126435.
- 48. Stroikova, V., Regul, D. & Meder, B. (2020). Rare Case of Selenite Poisoning Manifesting as Non-ST-segment Elevation Mycardial Infarction. *JACC: Case Reports*, 3(5), 811-815.
- 49. Kar, D., Sur, P., Mandal, S., Saha, T., & Kole, R. (2008). Assessment of Heavy Metal Pollution in Surface Water. *International Journal of Science and Technology*. 5, 119-124.
- 50. Mwegoha, W. J. S., & Kihampa, C. (2010). Heavy Metal Contamination in Agricultural Soils and Water in Dar es Salaam City, Tanzania. *African Journal Environmental Science and Technology*, 4(11)**,** 763-769.
- 51. Reza, R., & Singh, G. (2010). Heavy Metal Contamination and it's Indexing Approach for River Water. *International Journal of Environmental Science and Technology*, 7(4)**,** 785-792.
- 52. Nyakairu, W. A. G., & Christian, K. (2000). Mineralogical and Chemical Composition and Distribution of Rare Earth Elements in Clay-Rich Sediments from Central Uganda. Institute of Geochemistry, University of Vienna, Althanastrasse 14, A-1090 Vienna, Austria.
- 53. Fabjola, B., Marco, L., Laura, B., Alberto, B., Laura, E. D., & Elza, B. (2015). Evaluation of Heavy Metals Contamination from Environment to Food Matrix by TXRF: The Case of Rice and Rice Husk. *Journal of Chemistry*, 2015, 1-12.
- 54. Mangun, S. J. (2009). Microwave Digestion EPA Method 3052 on the Multiwave 3000. Field Application Report, PerkinElmer Inc, Shelton, CT 06484. USA.
- 55. Kingston, H. M. & Walter, P.J. (1998). The Art and Science of Microwave Sample Preparations for Trace and Ultratrace Elemental Analysis in Inductively Coupled Mass Spectrometry, A. Montaser, Wiley-VCH, New York. Pp 33-81.
- 56. Gathumbi, J. K., Kanja, L. W., Maitho, T. E., Mbaria, J. M., Nduhiu J. G., Gitau, F. K., Nderitu, J. G., Lucy, M. W. & Maloba, K. (2013). Assessment of Lead and Copper in Fish and Soil Sediments in Kirinyaga South District, Kenya. *Journal of Applied Sciences in Environmental Sanitation*, 8(3),145-150
- 57. Moywaywa, B. A. (2018). Heavy Metal Pollution Sediment, Water and Flora Along Thika River. Master of Science Thesis, University of Nairobi, Kenya. Pp 44-48.

https://ijasre.net/ Page 83

- 58. Omondi, C. (2017). Distribution of Heavy Metals in Water and Sediments of Lower River Nzoia, Master of Science Thesis, University of Nairobi, Kenya. Pp 30-36.
- 59. Idiriah, T. J. K., David-Omiema, S. & Ogbonna, D. N. (2012). Distribution of Heavy Metals in Water and Sediment Along Abonnema Shoreline, Nigeria. *Resources and Environment*, 2(1),33-40.
- 60. Wasike, P. W., Nawiri, M. P. & Wanyonyi, A. A. (2019). LEVELS of Heavy Metals (Pb, Mn, Cu & Cd) in Water in River Kuywa and Adjacent Wells. *Environment and Ecology Research*, 7(3), 135-138.
- 61. Muiruri, J. M., Nyambaka, J. M. & Nawiri, M. P. (2013). Heavy Metal in Water and Tilapia Fish from Athi-Galana-Sabaki Tributaries, Kenya. *International Food Research Journal*, 20(2), 891-896.
- 62. Water Services Regulatory Board (WASREB) (2008). Guidelines on Water Quality and Effluent Monitoring, Nairobi, Kenya. Pp 25-27.

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