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# Effect of Antioxidant activity of Astaxanthin in improving Human Spermatozoa lipid Peroxidation

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# Abstract

Malondialdehyde (MDA) is the final product of lipid peroxidation which reacts with Thiobarbituric Acid (TBA) to produce red color and can be observed by spectrophotometry at a wavelength of 532 nm. Astaxanthin is an antioxidant that can reduce MDA by counteracting free radicals in lipid peroxidation in human spermatozoa. The objective of this study was to identify the antioxidant activity of astaxanthin in improving human spermatozoa lipid peroxidation, with parameters of color, odor, pH, volume, viscosity, concentration, morphology, motility and MDA levels. In this study, astaxanthin was given to 19 patients orally in a single dose of 8 mg per day for 30 days. As a negative control, six people were given with CMC Na placebo. MDA level measurement was performed using a spectrophotometer and the quality of spermatozoa was measured using WHO 2010 parameters. Significant difference in MDA level was found before and after astxanthin administration with a p-value of 0.022 (p<0.05) and significant difference in sperm motility was also found with a p-value 0.000 (p<0.01), spermatozoa morphology with p 0.021 (p<0.05), and concentration with p 0.001 (p<0.05). There was no significant difference before and after astxanthin administration in sperm viscosity with p 0.414 (p>0.05), volume with p 0.965 (p>0.05) and pH with p 0.401 (p>0.05). In conclusion, astaxanthin can reduce lipid peroxidation, improve morphology, increase concentration and motility of human spermatozoa.

Keywords: Astaxanthin, lipid peroxidation, MDA, Human spermatozoa, Male infertility.

## **1. INTRODUCTION**

The prevalence of infertility throughout the world has increased over time and has reached around 15% [1,2], the contribution of male factors to infertility by 36.8% [3] and causes in idiopathic men by 31.7% [4]. Oxidative stress (OS) causes 30%-80% of cases of infertility in males by reducing spermatozoa motility and suppressing fertility potential [5,6,7]. OS occurs because of the imbalance due to excessive production of reactive oxygen species (ROS) so that it is unable to be neutralized by antioxidants in the spermatozoa cytoplasm [8] resulting in oxidative damage to membrane lipid, protein, and DNA [9]

Spermatozoa cell membrane contains large amounts of polyunsaturated fatty acids which are very easily oxidized by ROS. Free radicals will cause lipid peroxidation which will be formed in a longer chain. As a result, membrane permeability increases, and spermatozoa membrane integrity decreases [10, 11], 2011), cellular enzymes become inactive, and then the DNA structures will be damaged and and cell apoptosis will occur. As a result, the activity and number of sperm cells decreases, its motility decreases, producing sperm with abnormal morphology, and finally causes infertility [8]. The motility of spermatozoa is a factor that has a significant effect and is a major cause of infertility [12,13,14].

The measurement of the level of lipid peroxidation is performed by measuring the final product, the malondialdehyde (MDA), a dialdehyde compound as the end product of non-saturated fatty acid oxidation which is toxic to cell membranes [15]. Measurement of MDA levels is an indirect measurement of free radical activity as an indicator of oxidative stress which can be carried out by Thiobarbituric Acid Reactive Substances (TBARS test) with visible spectrophotometer methods [16]. In cases of idiopathic infertility it is recommended to perform ROS testing, one of which is by determining the MDA level [17].

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This lipid peroxidation can be prevented by fat-soluble antioxidants [18]. The negative effects of ROS on these cell membranes can be reduced by antioxidants. Astaxanthin is a fat-soluble antioxidant that functions as a radical scavenger so that it can prevent the initiation stage and contains peroxyl radicals so that it can prevent the propagation stage [19]. Astaxanthin is an antioxidant that has an effectiveness of 500-1000 times better than vitamin E in preventing fat peroxidation in vivo, 6000 times than vitamin C, 800 times than Coenzyme Q10, 560 times than green tea extract, and 75 times than lipoic acid [20]. Astaxanthin (3,3'-dihydroxy-4,4'-diketo-beta, beta'-carotene) has a radical group which has a strong activity of neutralizing free radicals and provides protection against lipid peroxidation and oxidation damage by cell membranes and tissues and has an anti-inflammatory effect by inhibiting cytokines [21, 22, 23].

## 2. MATERIALS AND METHODS

## 2.1. Materials

Distilled water (Brataco), Astaxanthin caplets 4 mg, 0.5% Eosin B in distilled water (Merck), 10% Negrosin solution in distilled water (Merck), 20% trikoloacetic acid, 1% Na Thiobarbiturate and MDA solution, HCl 1 N. The astaxanthin was in the form of 4 mg caplets derived from *Haematococus pluvialis* green algae produced by PT. X Jakarta. CMC Na 30 mg which is put in capsules was used as placebo.

#### 2.2. Instruments

Glass (Pyrex), Elliason pipette, centrifuge (Eppendorf 5403), multi-channel pipette (BioRad, USA), hemocytometer (Perkin Elmer 9800), pH meter (Eutech), votex (Maxi Mix II type 37,600), phase contrast microscope (Nikon), centrifuge (Eppendorf 5403), water bath, and HP 8452A Diode Array Spectrophotometer.

#### 2.3. Sample

This was an experimental study. Samples were obtained from one hospital in Surabaya after obtaining approval from the Health Research Ethics Committee, Dr. Soetomo Hospital, Surabaya, Indonesia with Ethical Clearance No. 710/Panke.KKE/XII/2016.

Subjects of this study were 25 persons, consisting of 19 persons in the sample group who were given with 8 mg astaxanthin per day in a single dose for 30 days and six persons in the placebo group who were given with CMC-Na 30 mg every day for 30 days. Everyone has signed informed consent about the treatment or examination that will be carried out after receiving enough information and approved without coercion by the concerned.

Sample of the study was sperm from subjects with primary or secondary infertility who came routinely for examination and had signed informed consent or statement of agreement regarding the treatment or examination that they will carry out after obtaining sufficient information and approved without coercion by the person concerned.

## 2.4. Sampling

Preceded by a period of sexual abstinence (no discharge of sperm fluid in any way) for 3-5 days, sperm retrieval was carried out in a separate room which was done by masturbation, and ejaculated into a wide-mouth sterile container. All ejaculate was stored and the temperature was maintained between  $20^{0}$ - $40^{0}$ C [24].

## 2.5. Sperm quality analysis

According to WHO 2010, it includes color, odor, pH, volume, viscosity, concentration, morphology and motility at the time before and after administration of the astaxanthin or placebo.

#### 2.6. Procedure for determining MDA levels

A total of 0.5 mL was added with 1.0 mL of 30% TCA, and centrifuged at 2500 rpm for 5 minutes. The supernatant was pipetted as much as 0.7 mL, added with 200  $\mu$ l of 1% Na Thiobarbiturate, then added with HCl 1 N up to 10.0 ml, then incubated for 135 minutes on the water bath. The absorption was observed by spectrophotometer method in a wavelength of 532 nm [5].

#### 2.7. The making of MDA standard solutions

MDA (50.4 mg) was mixed with distilled water to 100.0 mL (504.0  $\mu$ g/mL), pipetted as much as 1.0 ml, dissolved with distilled water to 10.0 mL (50.4  $\mu$ g/mL). MDA (50.4  $\mu$ g/mL) was pipetted as much as 6; 10; 12; 20; 30; 50; 70 and 125  $\mu$ L, then added with 1.0 mL of 30% TCA and 200  $\mu$ L of 1% Na Thiobarbiturate. The incubation was performed for 135 minutes on a water bath and its absorption was observed with spectrophotometer at a wavelength of 532 nm [5].

## **3. RESULTS AND DISCUSSION**

#### **3.1. Sample characteristics**

Of 25 persons, primary infertility was found in 72% and secondary infertility was in the remains. The average length of marriage was  $3.93\pm4.7000$  years, the number of children owned was  $0.41\pm0.7703$  children, the average age was  $25.48\pm6.6651$  years, under 40 years so they were not in the age of Advanced Paternal Age (APA) which has a significant impact on infertility [25]. The demographic profile of the sample characteristics is shown in Table 1.

Table 1. The sample characteristics							
Biodata	L	Values					
Age		25.48± 6.6651 years old					
Occupation:							
-	Entrepreneur	12(48%) persons					
-	Online driver	6(24%) persons					
-	Government employee	5(20%) persons					
-	No formal job	2(8%) persons					
Length of marriafe		3.93± 4.7000 years					
Number of children		0.41 ±0.7703 children					
Invertility status							
-	Primary invertility	18(72%) persons					
-	Secondari invertility	7(28%) persons					

#### 3.2. Color and odor of spermatozoa

The color and the odor w as normal at the time of, before, or after the administration of placebo and astaxanthin. The color of the sperm was grayish white, a distinctive odor of sperm like chlorine due to by sperm oxidation, an aliphatic polyamine that is produced by the prostate gland [26, 27].

#### 3.3. Spermatozoa pH

In this study, the pH was in the normal range, which was 7.2-7.8. There was no significant differences in either group, both before and after the administration of placebo and astaxanthin. Similar results were reported in studies conducted in the US [28], Norway [29], and Pakistan [30,31].

#### 3.4. Volume

In placebo group, the average volume before was  $1.88\pm0.373$  mL and after was  $2.65\pm0.725$  mL. There was no significant difference before and after the administration of CMC-Na (p>0.05). In sample group, the average volume before was  $2.58\pm1.094$  mL and after  $2.97\pm2.056$  mL. There was no significant difference before and after astaxanthin administration (p>0.05). A retrospective study in Brazil showed that sperm volume was the only parameter that decreased with age [32]. Volume of spermatozoa before and after treatment in placebo and sample is shown in figure 1.

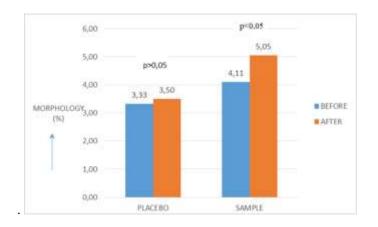


Figure 1. Volume of spermatozoa before and after treatment in placebo and sample.

## .3.5. Viscosity

In placebo group, the average viscosity before and after treatment was  $2.00\pm0$  cm. There were no significant differences before and after the administration of CMC-Na (p>0.05). In sample group, the average viscosity before treatment was  $2.26\pm0.733$  cm and

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after treatment  $2.16\pm0.375$  cm. There were no significant differences before and after astaxanthin administration (p>0.05). Viscosity was negatively correlated with motility with a correlation coefficient value of -0.045 (p>0.05). Spermatozoa viscosity before and after treatment in placebo and samples is shown in figure 2.

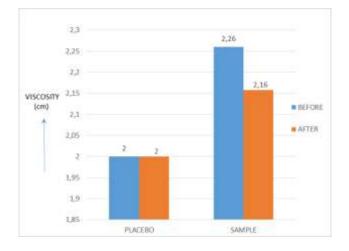


Figure 2. Spermatozoa viscosity before and after treatment in placebo and samples

## 3.6. Concentration

In placebo group, the average concentration before treatment was  $67.33\pm52.015 \times 10^6$  /mL and after treatment  $56.48\pm52.472 \times 10^6$  /mL. There were no significant difference before and after the administration of CMC-Na (p>0.05). In sample group, the average volume before treatment was  $34.29\pm22.383 \times 10^6$  /mL and after treatment  $41.84\pm18.610 \times 10^6$  /mL. There were significant differences before and after Astaxanthin administration (p>0.05). The concentration correlates with motility, and the correlation coefficient value was 0.003 (p<0.05).

Normal sperm concentration was 15 million sperm/mL or more [24]. The number or concentration of sperm in this study showed an increase but not significant. This was because there was no strict control to prevent disruption of spermatogenesis process that affects spermatozoa concentration, such as diet, smoking, alcohol, lifestyle, or the surrounding environment [33, 34].

Analysis of spermatozoa concentration showed a significant difference between before and after Astaxanthin administration. The lower the concentration of spermatozoa, the more infertile a man is, but rarely is there a man who does not produce sperm at all. This decrease in sperm concentration can be caused by disruption of spermatogenetic process due to, for example, high testicular temperature or smoking [27, 35]. ]. Spermatozoa concentration of before and after treatment in placebo & sample is shown in figure 3.

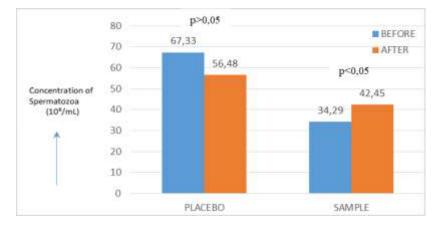


Figure 3. Spermatozoa concentration of before and after treatment in placebo & sample

## 3.7. Morphology

In placebo group, the average morphology before treatment was  $3.33\pm1.225\%$  and after treatment  $3.5\pm1.378\%$ . There were no significant differences before and after the administration of CMC-Na (p>0.05). In the sample group, the average morphology was

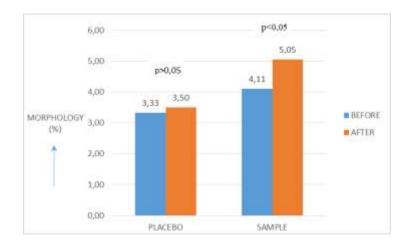
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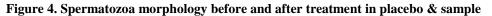
 $4.11\pm1.560$  and after treatment  $5.05\pm1.779\%$ . There were significant differences before and after Astaxanthin administration (p>0.05). Morphology correlates with motility with a correlation coefficient value of 0.000 (p<0.05).

The morphological changes in spermatozoa are affected by testicular function. The more normal the head, the better the function of the testes. Changes in the shape of the head and tail occur in the testes during spermatogenesis process. Nicotine endangers the process of spermatogenesis because it increases the production of cholesterol, triglycerides, phospholipids, free fatty acids in testicles and peroxidative damage [34, 36].

Changes in the shape of spermatozoa head affect sperm maturity, which also affects theotility and viability [27, 37]. There is a correlation between the normal morphology and pregnancy rates in vivo and in vitro [38].

Astaxanthin has a long structure that can increase stiffness and mechanical strength to protect the body from oxidation on cell membranes and it is hydrophilic and lipophilic so that it can more effectively inhibit reactive oxygen species against lipid peroxidation [20, 39, 40], so that it can improve the morphology of spermatozoa. Spermatozoa morphology before and after treatment in placebo & sample is shown in figure 4.





#### 3.8. Malondialdehyde (MDA)

In placebo group, the average concentration of MDA spermatozoa before treatment was  $2.1\pm2.379 \text{ nmol/10}^6$  /mL and after treatment  $1.65\pm1.537 \text{ nmol/10}^6$  /mL. There were no significant differences before and after CMC-Na administration with a p value 0.345 (p>0.05). In the sample group, the average concentration of MDA before treatment was  $1.74\pm1.146 \text{ nmol/10}^6$  /mL and after treatment  $1.27\pm0.672 \text{ nmol/10}^6$  /mL. There were significant differences before and after Astaxanthin administration with p 0.022 (p>0.05). In total group (placebo and sample) there were significant differences before and after treatment with p 0.009 (p<0.01). The MDA had negative correlation with motility with a correlation coefficient of 0.006 (p<0.05). ). Spermatozoa MDA before and after treatment in placebo and samples is shown in figure 5.

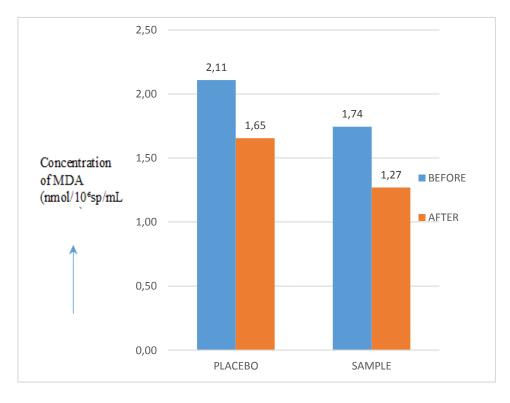


Figure 5. Spermatozoa MDA before and after treatment in placebo and samples

Receiver Operating Characteristic (ROC) analysis on MDA factor revealed AUC (Area Under Curve) value of 0.662 which was higher than 0.5. The cut-off value for the MDA factor was less than 1.98 with a sensitivity value of 100% and a specificity of 35.3%. This showed that if the MDA value is <1.98, the prediction of the motility level normality (>40) will be 100%. If the MDA value is >1.98, the prediction of the motility level will be included in abnormal category (<40) which is 35.5%. Figure 6 shows the results of ROC analysis on the MDA factor. MDA cut-off values (nmol/10<sup>6</sup>Spermatozoa/ml) on spermatozoa motility is shown in figure 6.

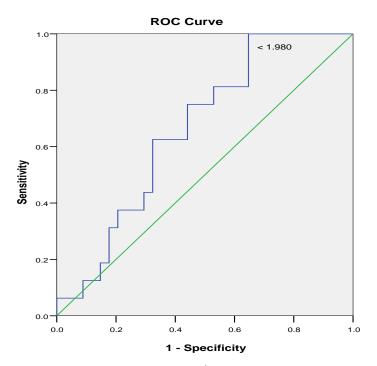


Figure 6. MDA cut-off values (nmol/10<sup>6</sup>Sp/ml) on spermatozoa motility

Spermatozoa have polyunsaturated fatty acids in the plasma membrane that are useful for membrane fluidity and membrane fusion such as acrosome reactions and spermatozoa-ovum interactions, but it causes spermatozoa to be very susceptible to free radical attack and lipid peroxidation [34] (Alahmar, 2019). Lipid peroxidation is a key mediator of ROS-induced sperm damage

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that causes infertility. The end result of lipid peroxidation is the breakdown of unsaturated fatty acid chains and MDA production [34, 41].

Sperm MDA level is negatively correlated with spermatozoa membrane integrity because high levels of MDA will reduce cell membrane integrity and increase spermatozoa damage resulting in a decrease in sperm quality. So, the higher the MDA level, the lower the percentage of normal integrity of spermatozoa membrane. As a result, the activity and number of sperm cells decreases, produces sperm with abnormal morphology, decreases motility, decreases sperm tail movement and interaction between sperm-ovum membranes in the fertilization process fails [8, 32, 41, 42]. In the cases of idiopathic infertility it is recommended to carry out ROS examination, one of which is by determining the MDA level [17].

Malonaldehyde analysis is an indirect analysis of free radicals and is a fairly easy analysis to determine the amount of free radicals formed. Direct analysis of free radicals is very difficult to do, because radical compounds are very unstable, electrophile and the reaction takes place so quickly that the measurement is very difficult when it is in the form of free radical compounds [43].

This lipid peroxidation can be prevented by fat-soluble antioxidants. Astaxanthin is a fat soluble antioxidant that functions as a radical scavenger so that it can prevent the initiation stage and it has peroxyl radicals so that it can prevent the propagation stage [7,18,19]. Like tocopherols, astaxanthin will transfer hydrogen atoms with single electrons to capture peroxyl lipids, thus preventing lipid peroxidation. Astaxanthin is a carotenoid group that is a chain breaking antioxidant because, during a peroxidation chain reaction, pyroxyl and alkoxyl radicals will be cut to prevent the propagation stage to produce antioxidant radicals that are not reactive [41, 44].

Astaxanthin is a pure antioxidant that can maintain the structure of the bilayer membrane by reducing 40% lipid hydroperoxide (LOOH), while non-polar carotenoids, such as lycopene and beta-carotene, show a strong pro-oxidant effect which increasing 85% LOOH. Therefore, astaxanthin is needed in cases of male infertility because it can maintain cell membrane integrity, regulation of spermatogenesis and spermatozoa motility [45,46,47,48].

Astaxanthin has a unique molecular structure that allows it to work inside and outside the cell membrane. This provides better protection than beta-carotene and vitamin C which can be positioned inside the lipid bilayer. This serves as a protection against oxidative damage by various mechanisms, such as singlet oxygen cooling; radical scavenging to prevent chain reactions; preservation of membrane structure by inhibiting lipid peroxidation, and improvement of immune system function and regulation of gene expression. As reported by various authors, astaxanthin inhibits lipid peroxidation in biological samples [21,34,41,49,50].

## 3.9. Motility

In placebo group, the average motility before treatment was  $24.17\pm11.143\%$  and after treatment  $28.33\pm6.831\%$ . There were no significant differences before and after CMC-Na administration (p>0.05). In sample group, the average motility before treatment was  $23.42\pm11.187$  and after treatment  $39.74\pm4.556\%$ . There were significant differences before and after Astaxanthin administration (p<0.05). Motility of spermatozoa before and after treatment on placebo and samples is shown in figure 7

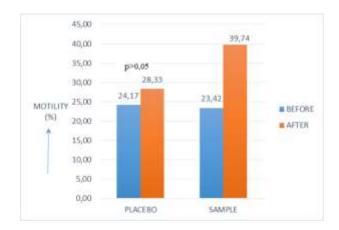


Figure 7. Motility of spermatozoa before and after treatment on placebo and samples

Spermatozoa motility requires energy, the ATP, which is mostly produced in the midpiece of the mitochondrial flagellum via the OXPHOS pathway [51,52]. During the OXPHOS process, 1-2% of oxygen consumed is released as reactive oxygen species (ROS), which can damage spermatozoa [53,54].

The decrease in spermatozoa motility is partly due to high ROS production, which is not able to be neutralized by the antioxidant defense system in spermatozoa or sperm plasma [9,55]. Spermatozoa plasma membrane is rich in unsaturated fatty acids, but has few antioxidant enzymes in the cytoplasma so they are very susceptible to oxidative damage, so membrane permeability increases and spermatozoa membrane integrity becomes low [11,2]. Disruption of membrane structure and function produces malondialdehyde compounds (MDA) which have a significant contribution to male infertility [2]. If motility is low, the migration of spermatozoa is not achieved, so the fertilization process does not occur, resulting in an infertile state. The Parameters of semen quality in the placebo and treatment groups is shown in Table 2.

Parameter	Placebo group		Treatment group			
	Before	After	Comparability	Before	After	Comparability
			test (a 0,05)			test (a 0,05)
Color	Normal	Normal	-	Normal	Normal	-
Odor	Normal	Normal	-	Normal	Normal	-
рН	7,2-7,8	7,2-7,8	-	7,2-7,8	7,2-7,8	-
Volume (mL)	$1.88 \pm$	$2.65 \pm$	p>0,05	$2.58 \pm$	$2.97 \pm 2.056$	p>0,05
	0.373	0.725		1.094		
Viscosity (cm)	$2.00\pm0.00$	$2.00\pm0.00$	p>0,05	$2.26 \pm$	$2.16\pm0.375$	p>0,05
				0.733		
Concentration	$67.33 \pm$	$56.48 \pm$	p>0,05	$34.29 \pm$	$41.84 \pm$	P<0,05
(x10 <sup>6</sup> /mL)	52.015	52.472		22.383	18.610	
Morphology (%)	3.33 ±	$3.5\pm1.378$	p>0,05	$4.11 \pm$	$5.05 \pm 1.779$	p<0,05
	1.225		_	1,560		-
Motility (%)	$24.17 \pm$	$28.33 \pm$	p>0,05	$23.42 \pm$	$39.74 \pm$	p<0,05
	11.143	6.831		11.187	4.556	

Table 2. Parameters of semen quality in the placebo and treatment groups

Antioxidant therapy is needed because seminal antioxidant level is unable to compensate for increased ROS production [56]. Astaxanthin is a natural carotenioid group needed to maintain cell membrane integrity and regulation of spermatogenesis that affects sperm motility and results in male infertility [46,57]. The effect of astaxanthin manifests through several mechanisms, such as electron scavenger through conjugated double bonds, and inhibition of lipid peroxidation. Recent research on the antioxidant activity of astaxanthin suggests that intraperitoneal injection of astaxanthin can increase sperm motility, amount, and viability, by reducing lipid peroxidation and increasing antioxidant defense [58].

Motility and MDA have a negative correlation with the correlation coefficient value of -0.356 and p value 0.006 (p<0.05). High MDA levels can result in decreased sperm motility [5]. High MDA level is influenced by lipid peroxidation level, which indirectly indicates high number of free radicals [15], which increases membrane permeability, cellular enzyme inactivation, damage to DNA structure and cell apoptosis, which results in decreased motility [8].

## 4. CONCLUSION

Astaxanthin in a single dose of 8 mg per day consumed for 30 days reduce lipid peroxidation and improve the concentration, morphology and motility of spermatozoa, so it can improve the function of human spermatozoa.

## ACKNOWLEDGMENT

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## **CONFLICT OF INTERESTS**

The authors declare that they have no conflict of interests in publication of this article.

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